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Association of Butyrophilin gene polymorphism (A465G) with milk production traits in Holstein Friesian crossbred cattle of Kerala

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Abstract

The butyrophilin (BTN1A1) gene is found at a quantitative trait locus (QTL) in cattle for milk production traits. The effect of a single nucleotide polymorphism (SNP) A465G in exon 8, which causes a lysine to arginine transition, on milk production attributes in Holstein Friesian crossbred cattle from Kerala was investigated in this study (151 numbers). Using custom synthesised primers, a 90-bp fragment encompassing the polymorphic region was amplified from genomic DNA isolated. Genotyping was carried out by high resolution melt curve analysis (HRM) and two genotypes KK (0.25) and KL (0.75) were detected based on melting temperature and melt curve patterns. Sanger's sequencing and sequence analysis of representative samples confirmed the genotypes. Chi-square test showed that the population was not distributed as per Hardy-Weinberg equilibrium ($p \le 0.05$). The relationship between the A465G transition and milk production traits like 305 day milk, fat, SNF yields, fat and SNF per cent was determined by general linear model-analysis of variance (GLM-ANOVA). In the model herd, season of calving, parity of animal (non-genetic factors), and genotype were considered fixed variables and milk production traits as dependent variable. The study revealed significantly higher ($p \le 0.01$) milk fat and SNF per cent for KK genotype (4.14±0.08 %; 7.86±0.06 %) than KL genotype (3.91±0.07 %; 7.73±0.05 %). The butyrophilin gene polymorphism (A465G) can be recommended as a marker for higher milk fat and SNF per cent in future breeding programmes in crossbred cattle of Kerala.

Keywords: Butyrophilin, SNP, high resolution melt curve, crossbred cattle, milk production traits

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Butvrophilin (BTN1A1) aene is mapped to bovine autosome 23 (Taylor et al., 1996) and in the same region quantitative trait loci (QTL) for milk production traits (Ashwell et al., 1996; Bennewitz et al., 2004). Previous researchers suggested BTN1A1 as a candidate gene for milk production (Komisarek et al., 2006; Weniun et al., 2009: Muszvnska et al., 2010: Al-Waith 2019) and disease resistance traits (Smith et al., 2010) in cattle. Butyrophilin gene is present on juxta-telomeric region of bovine leucocytic antigen (BoLA) complex (Ashwell et al., 1996) and span over 7003 bp length with eight exons and seven introns (Vishwanath, 2014). Butyrophilins are type I trans membrane proteins belonging to immunoglobulin (Ig) super family with an extra cellular and cytoplasmic domain. Cytoplasmic domain of BTN1A1 tightly binds to xanthine dehydrogenase (XDH) to form a supra-molecular complex. This complex binds to adipophilin in the phospholipid bilayer of milk secretory granule and pinches off the mammary gland epithelial cell in zipper like fashion (Robenek et al., 2006).

Many molecular markers in candidate genes for milk production such as Diacylglycerolo-transferase 1 (Lali and Anilkumar, 2016), Beta 1,4-galactosyltransferase-1 (Valsalan et al., 2021), Leptin (Lali and Bindu, 2015) and Osteopontin (Lali et al., 2020) were studied in crossbred cattle of Kerala. It is worth to note that, majority of SNPs act as indirect markers and the influence on production traits depends on state of linkage disequilibrium (LD) in cattle. Schmid and Bennewitz (2017) explained that the LD between the marker and the QTL might be affected by the genetic recombination during gamete formation. It reiterates the importance of association results of local populations to confirm the effects of markers especially indirect markers. The influence of BTN1A1gene polymorphisms was not studied yet in the crossbred cattle population of Kerala. The present study was designed to analyse the effect of polymorphism inBTN1A1 gene (A465G) on milk production traits in Holstein Friesian crossbred cattle of Kerala.

Materials and methods

Estimation of milk production traits

Milk samples were collected from 151 crossbred cattle maintained at University Livestock Farm and Fodder Research Development Scheme (ULF-FRDS), Mannuthy and cattle breeding farm (CBF), Thumburmuzhy once in a month for ten months. The samples were brought to laboratory under refrigerated conditions. Data regarding animal number. date of calving, parity, test day milk yield, recording month and date were obtained from farm records. Test day fat per cent was estimated from automatic milk analyser (MRC instruments) only after routine standardisation with Gerber's centrifugation method. Data regarding test day SNF per cent was derived from milk analyser by running samples. Test interval method (TIM), according to guidelines of international committee for animal recording (ICAR, 2020) was used to calculate 305 day vields of milk, fat and SNF along with 305 day milk fat and SNF per cent.

High resolution melt curve analysis

A volume of 5 mL blood was collected from jugular vein of 151 crossbred cattle in a EDTA coated vial. Genomic DNA was isolated from blood by standard phenol chloroform extraction method (Sambrook and Russell, 2001). Isolated DNA was checked for concentration and purity by Nanodrop spectrophotometry. Quality was assessed by 0.8 per cent agarose gel electrophoresis.

Polymorphism analysis and genotyping were carried out using High resolution melt curve (HRM) analysis (Desai et al., 2021) in Eco Real-Time PCR system (Illumina). Primers were custom synthesised using primer3 V.0.4.0 software and a gradient PCR was carried out to fix optimum annealing temperature and specific amplification was detected by running samples on 2.5 per cent agarose gel using molecular weight marker of 50 bp size. The HRM reaction was carried out using SSO FAST EVA GREEN super mix, forward (5'GCCCTTCTTCTGCTTGTGGT3') and reverse (5'TCAGCAACTACCATGACTCCC3') primers and template DNA. Thermal profile of reactions include 94°C for 5 min, 94°C for 30s, 62.5°C for 30s, 72°C for 30s followed by melt curve analysis. The results were confirmed

by Sanger's sequencing the representative samples from each genotype after detecting them using melting temperature (T_m) shift and melt curve pattern. Further, the genotype and allele frequencies were calculated and population was checked for Hardy Weinberg equilibrium using *chi*-square test.

Association with milk production traits

In order to study the association of A465G polymorphism with milk production traits, General linear model-Analysis of variance (GLM-ANOVA) was performed using SPSS version 24.0. Non-genetic factors such as herd, season of calving (October to January – post monsoon, February to May – summer, June to September – monsoon) and parity (1 to 4) and milk production traits 305-day milk yield, fat yield, fat per cent, SNF yield and SNF per cent as dependent variables.

The model was
$$Y_{ijklm} = \mu + H_i + S_i + P_{\kappa} + G_i + e_{iiklm}$$

Results and discussion

A single nucleotide polymorphism in the exon 8 of *BTN1A1*gene resulting from

adenine to guanine transition was studied in detail in the present study by high resolution melt curve analysis in HF crossbred cattle of Kerala.

High resolution melt curve analysis

Gradient PCR detected optimum annealing temperature of 62.5 °C. Specific amplification was confirmed by running amplicons through 2.5 per cent agarose gel (Fig. 1). Melt curve analysis clearly depicts presence of two curve patterns representing two genotypes KK and KL (Fig. 3) in the studied population. According to classification of Venter et al. (2001), the typical melting temperature shift of C/T or G/A mutations should be >0.5°C. Similarly, the genotypes of HRM analysis of the current study showed a melting temperature shift around 0.6°C. The chromatograms of both genotypes are depicted in Fig. 3. The sequence results were checked for any other genetic variations in the population since, multiple variations in the same amplicon will interfere the interpretation of HRM results.

The allele K (0.63 %) and genotype KL (0.75 %) were found to be frequent in the studied population (Table 1). T h e frequency of dominant allele K in different studies were0.88 by Komisarek and Dorynek (2003), 0.83 by Bhattacharya *et al.* (2006), 0.86 by Sadr *et al.* (2008), 0.88 by Rengarajan (2011), 0.89 by Vishwanath (2014) and 0.86 by Al-Waith (2019). Thus, the results of K

 Table 1. Genotype and allele frequencies for SNP A465G in bovine BTN1A1 gene

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SNP	Genotype frequencies			Allele frequencies		Chi-square value	
A465G (n=151)	KK/0.25 (37)	KL/0.75 (114)	LL/0 (0)	K/0.63	L/0.37	55.54 ^s	

S-Significant (p≤0.05)

Table 2. Effect of SNP A465G on milk production traits in crossbred cattle of Kerala

SI. No.	Trait	A	n volue	
	(Mean ± SE)	KK	KL	p- value
1.	305 day milk yield (kg)	2764.35±157.96	2850.68±128.20	p>0.05
2.	Fat yield (kg)	112.40±5.50	109.73±4.52	p>0.05
3.	Fat per cent	4.14±0.08	3.91±0.07	p≤0.01
4.	SNF yield (kg)	216.05±11.75	219.70±9.54	p>0.05
5.	SNF per cent	7.86±0.06	7.73±0.05	p≤0.01



Fig. 3. Normalised and difference melt curve of polymorphism A465G representing two different curve patterns for genotypes KK (blue) and KL (green), respectively.

allele dominance was in accordance with all aforementioned studies. The genotype LL was not detected in the present study which was in agreement with Sadr *et al.* (2008), Rychtarova *et al.* (2014), Vishwanath (2014) and Al-Waith (2019). *Chi*-square test showed

that the population was not in Hardy-Weinberg equilibrium ($p \le 0.05$). Natural selection may be going on favouring the K allele in crossbred cattle population. However, the status of HW equilibrium has to be further examined in a larger population.

Association with milk production traits

The two genotypes KK and KL of A465G transition of HF crossbreds of Kerala differed significantly (p≤0.01) with respect to fat and SNF per cent (Table 2) as per GLM ANOVA considering herd, season of calving, parity of animal (non-genetic factors) and genotype as fixed variables and milk production traits as dependent variables. Similar results were obtained by Rengarajan (2011) in milk fat per cent and current findings were also in accordance with Vishwanath (2014). However, contradictory to the present results, animals with KK genotype in a study by Komisarek et al. (2006) showed significant higher yields of milk, fat and SNF. Furthermore, Rychtarova et al. (2014) detected no association of the SNP A465G with milk production traits and it agrees with the current results except for fat and SNF per cent. Al-Waith (2019) identified significant difference between two genotypes KK and KL with respect to milk yield (KK had higher milk vield) and fat per cent (KL had higher milk fat per cent). Inconsistent association analysis results could be the result of population substructure creation, null alleles in population or excessive selection pressure (Lali et al., 2020). Thus, A465G polymorphism can be suggested as a potential marker for obtaining a good selling price to a farmer for milk as the fat and SNF per cent in milk determines the milk pricing in Kerala.

Conclusion

The study designed HRM analysis to genotype A465G transition in exon eight of bovine BTN1A1 gene. Genotype KK had significantly higher fat and SNF per cent in HF crossbreds of Kerala. The favourable allele K was found to be frequent in the population and it indicates that selection undergoes towards the favourable genotypes. Presently more emphasis is on milk composition traits and so extensive studies such as genome wide association studies are required to find out the genes regulating the milk components in cattle. Along with these, studies directing to find influence of stage of lactation are also necessary as it was found that this particular factor has effect on milk composition traits (Prasad and Subramanyam, 1986).

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Conflict of interest

The authors report no conflict of interest.

References

- Al-Waith, H. K. 2019. Association between BTN1A1gene polymorphism and milk production and its contents in Holstein cows. Plant Arch. 19: 1184-1187.
- Ashwell, M. S., Ogg, S. L. and Mather, I. H. 1996. The bovine butyrophilin gene maps to chromosome 23. Anim. Genet. 27: 171-173.
- Bennewitz, J., Reinsch, N., Paul, S., Looft, C., Kaupe, B., Weimann, C., Erhardt, G., Thaller, G., Kühn, C., Schwerin, M. and Thomsen, H., 2004. The DGAT1 K232A mutation is not solely responsible for the milk production quantitative trait locus on the bovine chromosome 14. J. Dairy Sci. 87(2): 431-442.
- Bhattacharya, T. K., Sheikh, F. D., Sukla, S., Kumar, P. and Sharma, A. 2007. Differences of ovine butyrophilin gene (exon 8) from its bovine and bubaline counterpart. Small Ruminant Res. 69: 198-202.
- Desai, A. G., Naicy, T., Aravindakshan, T. V., Muhasin, V. N. A., Bindu, L., Kurian, E., Bhat, V. and Akhil, G. H. 2021. Characterization of a 7 bp indel in MARCH1 promoter associated with reproductive traits in Malabari and Attappady Black goats of India. Small Ruminant Res. 204: 1-6.
- ICAR (International committee for animal recording) guidelines. 2020. Computing

of accumulated lactation yield, The Global Standard for Livestock Data, 4p.

- Komisarek, J. and Dorynek, Z. 2003. Polymorphism of BTN and GHR genes and its impact on bulls' breeding value for milk production traits. *J. Anim. Feed Sci.* **12**(4): 681-688.
- Komisarek, J., Waskowicz, K. and Dorynek, Z. 2006.Analysisoftherelationshipbetween two single nucleotide polymorphisms of the butyrophilin (*BTN1A1*) gene and milk production traits in Jersey cattle. *Ann. Anim. Sci.* **6**: 45-52.
- Lali, F.A. and Anilkumar, K. 2016. Diacylglycerolo-transferase 1 (*DGAT1*) gene and milk production traits in crossbred cattle of Kerala. *J. Vet. Anim. Sci.* **47** (1): 52-57.
- Lali, F.A., Anilkumar, K. and Aravindakshan, T. 2020. Effect of Osteopontin gene variants on milk production traits in Holstein Friesian crossbred cattle of Kerala. *Turkish J. Vet. Anim. Sci.* 44 (3): 695-701.
- Lali, F.A. and Bindu, K.A., 2015. Association of leptin gene polymorphism and milk yield in cattle. *Indian Vet. J.* **92** (1), pp.55-57.
- Muszynska, M., Szatkowska, I., Grzesiak, W., Dybus, A. and Zaborski, D. 2010. Two single nucleotide polymorphisms within bovine butyrophilin gene (BTN/HaeIII and BTN/SchI) and their association with milk performance traits in Jersey cattle. *Arch. Tierz.* **5**: 501-509.
- Prasad, V. and Subramanyam, M. 1986. Composition of milk of crossbred cattle. *J. Vet. Anim. Sci.* **17**: 33-45.
- Rengarajan, K.2011. Molecular characterization of bovine butyrophilin (*BTN1A1*) alleles in exon 3 and 8 and their association with fat content and SCC in milk of dairy cattle. *PhD Thesis*, National Dairy Research Institute, Karnal, 85p.

Robenek, H., Hofnagel, O., Buers, I., Lorkowski, S., Schnoor, M., Robenek, M. J., Heid, H., Troyer, D. and Severs, N. J. 2006. Butyrophilin controls milk fat globule secretion. *Proceeding of the national academy of sciences*. **103**: 10385-10390.

- Rychtarova, J., Sztankoova, Z., Kysclova, J., Stipkova, M., Vacek, M. and Stolc, L. 2014. Effect of *DGAT1*, *BTN1A1*, *OLR1* and *STAT1* genes on milk production and reproduction traits in the Czech Fleckvieh breed. *Czech J. Anim. Sci.* **59** (2): 45-53.
- Sadr, A. S., Nasiri, M. T. B., Alami-Saeid, K., Fayazi, J., Roshanfekr, H. and Mohammadi, M. 2008. DNA polymorphism of butyrophilin gene by PCR-RFLP technique. *Afr. J. Biotech.* **7**: 2527-2529.
- Sambrook, J. and Russell, D.W. 2001. *Molecular cloning: A laboratory manual.* (3rd Ed), Cold Spring Harbor Laboratory Press, New York. 1886p.
- Schmid, M. and Bennewitz, J. 2017. Invited review: Genome-wide association analysis for quantitative traits in livestock-a selective review of statistical models and experimental designs. *Arch. Anim. Breed.* **60**(3): 335-346.
- Smith, I. A., Knezevic, B. R., Ammann, J. U., Rhodes, D. A., Aw, D., Palmer, D. B., Mather, I. H. and Trowsdale, J. 2010. BTN1A1, the mammary gland butyrophilin, and BTN2A2 are both inhibitors of T cell activation. *J. Immunol.* 184: 3514-3525.
- Taylor, C., Everest, M. and Smith, C. 1996. Restriction fragment length polymorphism in amplification products of the bovine butyrophilin gene: assignment of bovine butyrophilin to bovine chromosome 23. *Anim. Genet.* 27: 183-185.
- Venter, J.C., Adams, M.D., Myers, E.W., Li, P.W., Mural, R.J., Sutton, G.G., Smith, H.O., Yandell, M., Evans, C.A., Holt, R.A. and Gocayne, J.D. 2001. The sequence

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of the human genome. *Science*. **291**: 1304-1351.

- Valsalan, J., Sadan, T., Venkatachalapathy, T., Anilkumar, K. and Aravindakshan, T.V. 2020. Identification of novel singlenucleotide polymorphism at exon1 and 2 region of *B4GALT1* gene and its association with milk production traits in crossbred cattle of Kerala, India. *Anim. Biotech*: 1-9.
- Vishwanath, B. 2014. Molecular genetic studies on butyrophilin gene in Malnad Gidda cattle and its association with milk quality traits. *M.V.Sc. Thesis*, Karnataka Veterinary, Animal and Fisheries Sciences University, Bidar, 149p.
- Wenjun, W., Keltui, O., Sheng, X. M., Chen, S. X. and Qin, C. 2009. Association of a polymorphism in exon 7 of butyrophilin gene with milk performance traits in dairy cow. *Acta Agric. Univ. Jiangxiensis*, **31**: 18-21.
- Yardibi, H., Gürsel, F.E., Ates, A., Akıs, I., Hosturk, G.T. and Oztabak, K. 2013. BTN1A1, FABP3 and TG genes polymorphism in East Anatolian red cattle breed and South Anatolian red cattle breed. *Afr. J. Biotech.* **12**(20): 2802-2807.
- Zegeye, A., Ashwell, M., Ogg, S., Rexroad, C. and Mather, I. H. 1999. RFLP markers in bovine butyrophilin gene. *Anim. Genet.* **30**: 385-386.